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# Prevalence of *Klebsiella Pneumoniae*, *Pseudomonas Aeruginosa* and *Staphylococcus Aureus* Isolated from Pus & Wound Swab & their Antibiotic Resistance Pattern

Lina Das<sup>1</sup>, Biswajit Batabyal<sup>2</sup>

<sup>1</sup>Sister Nivedita University, Kolkata, West Bengal, India

<sup>2</sup>Consultant Microbiologist; Serum Analysis Center Pvt. Ltd.; Howrah, West Bengal, India

**Abstract--** Studies on pus and wound swabs frequently identify *Staphylococcus aureus*, *Pseudomonas aeruginosa*, and *Klebsiella pneumoniae* as primary pathogens, with high rates of multidrug resistance (MDR) to common antibiotics. *Staphylococcus aureus* often shows significant methicillin resistance (MRSA) or Oxacillin resistance (ORSA) while *Pseudomonas aeruginosa* and *Klebsiella pneumoniae* exhibit high resistance to third-generation cephalosporins and, increasingly, carbapenems..

The presence of pathogenic bacteria in the wound does not imply infection. Infection occurs when one or more than one contaminant evades the host defenses, replicating in large numbers, attacks, and harms the host tissue. Different microbial organisms can infect wounds. They are likely *Pseudomonas aeruginosa*, *Staphylococcus aureus*, *Klebsiella pneumoniae*.

This study was included 220 cases of pus and swab from the out and in patients. This study was conducted in Serum Analysis Center Pvt. Ltd.; Howrah. The duration of study period of January, 2026 to March, 2026.

**Result--** In this study *Klebsiella pneumoniae* in all age group shows Resistant in Cephalosporins group, Fluoroquinolones group, Penicillin group. Highly sensitive rate of Antibiotics is Tigecycline. Colistin highly Resistant antibiotics in *Klebsiella pneumoniae* and *Pseudomonas aeruginosa* in all age group. Piperacillin/Tazobactam and Cefoperazone/Sulbactam both are good response in *Pseudomonas aeruginosa* in all age group. Cefoperazone/Sulbactam also good performance in *Klebsiella pneumoniae* in all age group. In our study in all age group approx. 48.1% Oxacillin Resistant (ORSA) *Staphylococcus aureus* are found. This findings is very important for our this clinical study. Linezolid and Tigecycline both are highly sensitive in *Staphylococcus aureus* in all age group.

This study concludes early detection and appropriate antibiotic application remain a significant priority in controlling the development and spread of multidrug resistant organisms.

**Keywords--** Pus/Wound specimen; *Klebsiella pneumoniae*; *Pseudomonas aeruginosa*; *Staphylococcus aureus*; Antibiogram; Resistance; ORSA.

## I. INTRODUCTION

Pus and wound infections are frequently caused by *Klebsiella pneumoniae*, *Pseudomonas aeruginosa*, and *Staphylococcus aureus*, which are leading causes of nosocomial infections. These pathogens exhibit high prevalence and significant multidrug resistance (MDR), including Methicillin-resistant *Staphylococcus aureus* (MRSA) or Oxacillin -resistant *Staphylococcus aureus* (ORSA) and extended-spectrum beta-lactamase (ESBL)-producing *Klebsiella pneumoniae*. High resistance to commonly used antibiotics like cephalosporins and Fluoroquinolones group necessitates urgent surveillance and tailored antibiogram-guided therapy.

Healthcare-associated infections (HAIs) represent a critical public health concern worldwide, contributing significantly to patient morbidity, mortality, and healthcare costs [1]. Among the various pathogens responsible for HAIs, *Staphylococcus aureus*, *Klebsiella pneumoniae* and *Pseudomonas aeruginosa* stand out as predominant causative agents, particularly in intensive care units where critically ill patients are at heightened risk [2]. The emergence and proliferation of antibiotic-resistant strains of these organisms have transformed the landscape of clinical microbiology and infection management. *Staphylococcus aureus*, a gram-positive bacterium, has demonstrated remarkable adaptability in developing resistance mechanisms against various antimicrobial agents. The evolution from methicillin-sensitive *Staphylococcus aureus* (MSSA) or Oxacillin sensitive *Staphylococcus aureus* (OSSA) to methicillin-resistant *Staphylococcus aureus* (MRSA) or Oxacillin resistant *Staphylococcus aureus* (ORSA) marked a significant milestone in the history of antibiotic resistance [3]. Currently, MRSA/ORSA infections pose substantial therapeutic challenges, with some strains exhibiting resistance to multiple drug classes, including glycopeptides, lincosamides, and fluoroquinolones [4].



The organism's ability to colonize both healthcare workers and environmental surfaces creates persistent reservoirs that facilitate transmission within healthcare facilities.

*Klebsiella pneumoniae*, a gram-negative member of the Enterobacteriaceae family, has emerged as a formidable pathogen in hospital settings. The widespread dissemination of extended-spectrum beta-lactamase (ESBL) producing *Klebsiella pneumoniae* strains has severely limited therapeutic options for treating infections caused by this organism [5]. Furthermore, the emergence of carbapenem-resistant *Klebsiella pneumoniae* (CRKP) has created scenarios where few or no effective antibiotics remain available for treatment [6]. This organism's propensity for horizontal gene transfer contributes to the rapid spread of resistance determinants among bacterial populations. The correlation between environmental contamination and clinical infections has gained increasing recognition in recent years. Healthcare environments, including medical equipment, surfaces, and the hands of healthcare workers, serve as potential reservoirs for resistant organisms [7]. Studies have demonstrated that environmental isolates often harbor similar resistance patterns to those found in clinical specimens, suggesting possible transmission pathways within healthcare facilities [8]. Understanding these correlations is essential for developing effective infection prevention and control strategies.

The intensive care unit environment presents unique challenges for infection control due to the high acuity of patients, frequent invasive procedures, and extensive use of broad-spectrum antibiotics. These factors create selective pressure that favors the survival and proliferation of resistant organisms [9]. Healthcare workers' hands, despite being recognized as the primary vehicle for pathogen transmission, continue to be implicated in the spread of multidrug-resistant organisms when proper hand hygiene practices are not consistently followed [10].

Current surveillance programs focusing on antibiotic resistance patterns provide valuable insights into the epidemiology of resistant organisms within healthcare settings. However, comprehensive studies examining the relationship between clinical isolates, environmental contamination, and healthcare worker colonization remain limited [11]. Such investigations are crucial for understanding transmission dynamics and implementing targeted interventions to reduce the burden of healthcare-associated infections.

The present study was designed to investigate the antibiotic resistance patterns of *Staphylococcus aureus* and *Klebsiella pneumoniae* isolated from various sources within a surgical intensive care unit, including clinical specimens, environmental samples, and healthcare workers' hands. By examining these correlations, we aim to provide insights that can inform evidence-based infection control practices and contribute to the development of comprehensive strategies for combating antibiotic resistance in healthcare settings.

Centers for Disease Control and Prevention (CDC) define surgical site infections (SSIs) as infections that occur at the site of incision within 30 days of any surgery. Despite of having advance techniques in surgery SSIs is become predominant reason for hospital-acquired infection, which causes mortality, Morbidity and increase in medical expenses.[12] Uncontrolled, spreading of antibacterial resistance among bacterial agents creates more challenge in clinical and surgical practice for management to treat surgical site infections. Increasing resistance in microbes directly affected the effectiveness of antimicrobials and causes worldwide problem. In developing countries, the surgical site infection and wound infection is more serious due to the irrational prescription of antibacterial agents.[13] *Pseudomonas aeruginosa* is the Gram-negative bacteria that have nearly replaced Gram-positive bacteria *Staphylococcus aureus* in hospital acquired infections with Gram-negative rods, which become more common in recent years. When compared to a decade long retrospective investigation *Pseudomonas aeruginosa* has an increased prevalence of wound infections. [14]

In this study is the prevalence and antibiotic resistance patterns of *Klebsiella pneumoniae*, *Pseudomonas aeruginosa*, and *Staphylococcus aureus* in pus and wound swabs are to identify the predominant pathogens causing wound infections, determine their susceptibility to commonly used antibiotics, and detect multidrug-resistant (MDR) strains.

## II. MATERIALS AND METHODS

### *Study Area:*

The present retrospective study was carried out in the Referral Laboratory, Serum Analysis Centre Pvt. Ltd.; 177, Netaji Subhas Road, Halder Para, Howrah-711101, West Bengal, India.



*Study Period:*

This study was carried out a period of Three months from January 2026 to March 2026.

*Study Samples:*

The inclusion criteria for this study in all patients of both sexes of outpatients of pus/ wound specimen.

*Sample Processing:*

The study aimed to analyze of *Klebsiella pneumoniae*, *Pseudomonas aeruginosa* & *Staphylococcus aureus* of pus & wound specimen for three months. These samples were transported to the microbiology laboratory following standard protocols to maintain integrity and prevent contamination ensuring prompt processing for accurate isolation and identification. Clinical specimens were streaked onto appropriate culture media such as blood agar and MacConkey agar.

The cultures were incubated aerobically at 37°C for overnight incubation. Bacterial colonies were examined for characteristic morphology, biochemical reaction and confirmed as *Staphylococcus aureus*, *Pseudomonas aeruginosa* and *Staphylococcus aureus* using the VITEK2 system (bioMérieux, Marcy-l'Étoile, France). The antimicrobial susceptibility of NFGNB isolates was determined according to established guidelines using the VITEK2 system. The VITEK2 system utilizes standardized antimicrobial susceptibility panels to assess susceptibility profiles against various antibiotics through automated turbidimetric methods. Susceptibility profiles were classified as susceptible, intermediate, or resistant. Data from the VITEK2 system, including identification and

antimicrobial susceptibility profiles, were exported and analyzed using Microsoft Excel.

To maintain the quality of the VITEK system, the instrument undergoes regular calibration following the manufacturer's instructions. The reagents, including microbial identification and susceptibility cards, are also routinely calibrated using control strains such as *Ps. aeruginosa* ATCC 27853, *Klebsiella pneumoniae* ATCC 13883, and *Staphylococcus aureus* *Staphylococcus aureus* ATCC 25923 (Oxacillin susceptible) and *Staphylococcus aureus* ATCC 43300 (Oxacillin resistant) were used as the control strains. The prevalence of multi-drug resistance among isolates was determined based on criteria for defining resistance to multiple antibiotic classes [15].

*Identification Of Isolates:*

The isolates were identified using colony morphology, Gram staining, Motility test, Indole test, Citrate test [Simmons Citrate Agar media], Urease test [Urease Agar media + 40% Urea], Triple Sugar Iron Agar media, ONPG [Ortho-nitrophenyl beta-D-galactopyranoside] , Oxidase test, Catalase test, coagulase test , D Nase test and ORSA /MRSA test [16, 17].

### III. RESULTS

*Commonly Prescribed Antibiotic Classes:*

1. Children (<18 years)
2. Adults (>18-64 years)
3. Older Adults (>64 years)

TOTAL NUMBER OF SAMPLE COLLECTION: 220

**Table: 1.: Distribution of Pathogens from Wound/Pus specimen:**

Pathogens	Total No. of Isolates	Total No. of Isolates	Total No. of Isolates
	[<18 Years]	[>18-64 Years]	[>64 Years]
<i>Klebsiella pneumoniae</i>	10	48	26
<i>Pseudomonas aeruginosa</i>	02	16	06
<i>Staphylococcus aureus</i>	16	38	06

**Table: 2.: Prevalence of pathogens isolated on PUS/WOUND SWAB culture:**

Pathogens	<18 Years		>18-64 Years		>64 Years	
	Male	Female	Male	Female	Male	Female
<i>Klebsiella pneumoniae</i>	10	00	30	18	16	10
<i>Pseudomonas aeruginosa</i>	02	00	12	04	03	03
<i>Staphylococcus aureus</i>	14	02	30	08	04	02

**Table: 3. Percentage of Resistant & Susceptibility of isolated *Klebsiella pneumoniae* to tested use of antibiotics in PUS/WOUND Culture [<18 Years]:**

**Total Isolates: 10**

Antibiotics	R (No.)	R (%)	S (No.)	S (%)
Amoxicillin/ Clavulanic Acid	06	60.0%	04	40.0%
Piperacillin+Tazobactam	06	60.0%	04	40.0%
Cefuroxime	08	80.0%	02	20.0%
Ceuroxime Axetil	09	90.0%	01	10.0%
Ceftriaxome	09	90.0%	01	10.0%
Cefoperazone/Sulbactam	02	20.0%	08	80.0%
Cefepime	08	80.0%	02	20.0%
Ertapenem	08	80.0%	02	20.0%
Imipenem	02	20.0%	08	80.0%
Meropenem	02	20.0%	08	80.0%
Co-trimoxazole [Trimethoprim+ Sulphamethoxale]	01	10.0%	09	90.0%
Amikacin	02	20.0%	08	80.0%
Gentamicin	02	20.0%	08	80.0%
Colistin	09	90.0%	01	10.0%
Tigecycline	01	10.0%	09	90.0%
Ciprofloxacin	08	80.0%	02	20.0%
Fosfomycin	04	40.0%	06	60.0%

**Table: 4. Percentage of Resistant & Susceptibility of isolated *Klebsiella pneumoniae* to tested use of antibiotics in PUS/WOUND Culture [>18-64 Years]:**

**Total Isolates: 48**

<b>Antibiotics</b>	<b>R (No.)</b>	<b>R (%)</b>	<b>S (No.)</b>	<b>S (%)</b>
<b>Amoxicillin/ Clavulanic Acid</b>	<b>42</b>	<b>87.5%</b>	<b>06</b>	<b>12.5%</b>
<b>Piperacillin+Tazobactam</b>	<b>34</b>	<b>70.8%</b>	<b>14</b>	<b>29.2%</b>
<b>Cefuroxime</b>	<b>38</b>	<b>79.16%</b>	<b>10</b>	<b>20.84%</b>
<b>Ceuroxime Axetil</b>	<b>36</b>	<b>75.00%</b>	<b>12</b>	<b>25.00%</b>
<b>Ceftriaxome</b>	<b>36</b>	<b>75.00%</b>	<b>12</b>	<b>25.00%</b>
<b>Cefoperazone/Sulbactam</b>	<b>26</b>	<b>54.10%</b>	<b>22</b>	<b>45.90%</b>
<b>Cefepime</b>	<b>26</b>	<b>54.10%</b>	<b>22</b>	<b>45.90%</b>
<b>Ertapenem</b>	<b>26</b>	<b>54.10%</b>	<b>22</b>	<b>45.90%</b>
<b>Imipenem</b>	<b>30</b>	<b>62.50%</b>	<b>18</b>	<b>37.50%</b>
<b>Meropenem</b>	<b>28</b>	<b>58.30%</b>	<b>20</b>	<b>41.70%</b>
<b>Co-trimoxazole [Trimethoprim+ Sulphamethoxale]</b>	<b>22</b>	<b>45.90%</b>	<b>26</b>	<b>54.10%</b>
<b>Amikacin</b>	<b>26</b>	<b>54.10%</b>	<b>22</b>	<b>45.90%</b>
<b>Gentamicin</b>	<b>24</b>	<b>50.00%</b>	<b>24</b>	<b>50.00%</b>
<b>Colistin</b>	<b>30</b>	<b>62.50%</b>	<b>18</b>	<b>37.50%</b>
<b>Tigecycline</b>	<b>08</b>	<b>16.60%</b>	<b>40</b>	<b>83.40%</b>
<b>Ciprofloxacin</b>	<b>38</b>	<b>79.10%</b>	<b>10</b>	<b>20.90%</b>
<b>Fosfomycin</b>	<b>08</b>	<b>16.60%</b>	<b>40</b>	<b>83.40%</b>

**Table: 5. Percentage of Resistant & Susceptibility of isolated *Klebsiella pneumoniae* to tested use of antibiotics in PUS/WOUND Culture [>64 Years]:**

**Total Isolates: 26**

<b>Antibiotics</b>	<b>R (No.)</b>	<b>R (%)</b>	<b>S (No.)</b>	<b>S (%)</b>
<b>Amoxicillin/ Clavulanic Acid</b>	<b>08</b>	<b>30.70%</b>	<b>18</b>	<b>69.30%</b>
<b>Piperacillin+Tazobactam</b>	<b>08</b>	<b>30.70%</b>	<b>18</b>	<b>69.30%</b>
<b>Cefuroxime</b>	<b>08</b>	<b>30.70%</b>	<b>18</b>	<b>69.30%</b>
<b>Cefuroxime Axetil</b>	<b>08</b>	<b>30.70%</b>	<b>18</b>	<b>69.30%</b>
<b>Ceftriaxome</b>	<b>08</b>	<b>30.70%</b>	<b>18</b>	<b>69.30%</b>
<b>Cefoperazone/Sulbactam</b>	<b>06</b>	<b>23.80%</b>	<b>20</b>	<b>76.20%</b>
<b>Cefepime</b>	<b>10</b>	<b>38.40%</b>	<b>16</b>	<b>61.60%</b>
<b>Ertapenem</b>	<b>10</b>	<b>38.40%</b>	<b>16</b>	<b>61.60%</b>
<b>Imipenem</b>	<b>08</b>	<b>30.70%</b>	<b>18</b>	<b>69.30%</b>
<b>Meropenem</b>	<b>04</b>	<b>15.40%</b>	<b>22</b>	<b>84.60%</b>
<b>Co-trimoxazole [Trimethoprim+ Sulphamethoxale]</b>	<b>04</b>	<b>15.40%</b>	<b>22</b>	<b>84.60%</b>
<b>Amikacin</b>	<b>08</b>	<b>30.70%</b>	<b>18</b>	<b>69.30%</b>
<b>Gentamicin</b>	<b>04</b>	<b>15.40%</b>	<b>22</b>	<b>84.60%</b>
<b>Colistin</b>	<b>22</b>	<b>84.60%</b>	<b>04</b>	<b>84.60%</b>
<b>Tigecycline</b>	<b>01</b>	<b>3.84%</b>	<b>25</b>	<b>96.16%</b>

**Table: 6. Percentage of Resistant & Susceptibility of isolated *Staphylococcus aureus* to tested use of antibiotics in PUS/WOUND Culture [<18 Years]:**

**Total Isolates: 16**

<b>Antibiotics</b>	<b>R (No.)</b>	<b>R (%)</b>	<b>S (No.)</b>	<b>S (%)</b>
<b>Cefoxitin Screen</b>	<b>10</b>	<b>62.50%</b>	<b>06</b>	<b>37.50%</b>
<b>Benzylpenicillin</b>	<b>15</b>	<b>93.75%</b>	<b>01</b>	<b>6.25%</b>
<b>Oxacillin</b>	<b>04</b>	<b>25.00%</b>	<b>12</b>	<b>75.00%</b>
<b>Gentamicin</b>	<b>01</b>	<b>6.25%</b>	<b>15</b>	<b>93.75%</b>
<b>Ciprofloxacin</b>	<b>15</b>	<b>93.75%</b>	<b>01</b>	<b>6.25%</b>
<b>Levofloxacin</b>	<b>15</b>	<b>93.75%</b>	<b>01</b>	<b>6.25%</b>
<b>Inducible Clindamycin Resistance</b>	<b>10</b>	<b>62.50%</b>	<b>06</b>	<b>37.50%</b>
<b>Erythromycin</b>	<b>10</b>	<b>62.50%</b>	<b>06</b>	<b>37.50%</b>
<b>Cotrimoxazole</b>	<b>01</b>	<b>6.25%</b>	<b>15</b>	<b>93.75%</b>
<b>[Trimethoprim+ Sulphamethoxale]</b>				
<b>Clindamycin</b>	<b>06</b>	<b>37.50%</b>	<b>10</b>	<b>62.50%</b>
<b>Linezolid</b>	<b>01</b>	<b>6.25%</b>	<b>15</b>	<b>93.75%</b>
<b>Daptomycin</b>	<b>01</b>	<b>6.25%</b>	<b>15</b>	<b>93.75%</b>
<b>Teicoplanin</b>	<b>01</b>	<b>6.25%</b>	<b>15</b>	<b>93.75%</b>
<b>Vancomycin</b>	<b>01</b>	<b>6.25%</b>	<b>15</b>	<b>93.75%</b>
<b>Tetracycline</b>	<b>01</b>	<b>6.25%</b>	<b>15</b>	<b>93.75%</b>
<b>Tigecycline</b>	<b>01</b>	<b>6.25%</b>	<b>15</b>	<b>93.75%</b>
<b>Nitrofurantoin</b>	<b>01</b>	<b>6.25%</b>	<b>15</b>	<b>93.75%</b>
<b>Rifampicin</b>	<b>01</b>	<b>6.25%</b>	<b>15</b>	<b>93.75%</b>

**Table: 7. Percentage of Resistant & Susceptibility of isolated *Staphylococcus aureus* to tested use of antibiotics in PUS/WOUND Culture [ >18-64 Years]:**

**Total Isolates: 38**

<b>Antibiotics</b>	<b>R (No.)</b>	<b>R (%)</b>	<b>S (No.)</b>	<b>S (%)</b>
<b>Cefoxitin Screen</b>	<b>16</b>	<b>42.10%</b>	<b>22</b>	<b>57.90%</b>
<b>Benzylpenicillin</b>	<b>26</b>	<b>68.40%</b>	<b>12</b>	<b>31.60%</b>
<b>Oxacillin</b>	<b>20</b>	<b>52.60%</b>	<b>18</b>	<b>47.40%</b>
<b>Gentamicin</b>	<b>18</b>	<b>47.40%</b>	<b>20</b>	<b>52.60%</b>
<b>Ciprofloxacin</b>	<b>36</b>	<b>94.70%</b>	<b>02</b>	<b>5.30%</b>
<b>Levofloxacin</b>	<b>26</b>	<b>68.40%</b>	<b>12</b>	<b>31.60%</b>
<b>Inducible Clindamycin Resistance</b>	<b>20</b>	<b>52.60%</b>	<b>18</b>	<b>47.40%</b>
<b>Erythromycin</b>	<b>26</b>	<b>68.40%</b>	<b>12</b>	<b>31.60%</b>
<b>Co-trimoxazole</b> <b>[Trimethoprim+ Sulphamethoxale]</b>	<b>08</b>	<b>21.10%</b>	<b>30</b>	<b>78.90%</b>
<b>Clindamycin</b>	<b>26</b>	<b>68.40%</b>	<b>12</b>	<b>31.60%</b>
<b>Linezolid</b>	<b>01</b>	<b>2.63%</b>	<b>37</b>	<b>97.37%</b>
<b>Daptomycin</b>	<b>06</b>	<b>15.7%</b>	<b>32</b>	<b>84.3%</b>
<b>Teicoplanin</b>	<b>04</b>	<b>10.5%</b>	<b>34</b>	<b>89.5%</b>
<b>Vancomycin</b>	<b>04</b>	<b>10.5%</b>	<b>34</b>	<b>89.5%</b>
<b>Tetracycline</b>	<b>06</b>	<b>15.7%</b>	<b>32</b>	<b>84.3%</b>
<b>Tigecycline</b>	<b>01</b>	<b>2.63%</b>	<b>37</b>	<b>97.37%</b>
<b>Nitrofurantoin</b>	<b>01</b>	<b>2.63%</b>	<b>37</b>	<b>97.37%</b>
<b>Rifampicin</b>	<b>01</b>	<b>2.63%</b>	<b>37</b>	<b>97.37%</b>

**Table: 8. Percentage of Resistant & Susceptibility of isolated *Staphylococcus aureus* to tested use of antibiotics in PUS/WOUND Culture [>64 Years]:**

**Total Isolates: 06**

<b>Antibiotics</b>	<b>R (No.)</b>	<b>R (%)</b>	<b>S (No.)</b>	<b>S (%)</b>
<b>Cefoxitin Screen</b>	<b>02</b>	<b>33.30%</b>	<b>04</b>	<b>66.70%</b>
<b>Benzylpenicillin</b>	<b>05</b>	<b>83.33%%</b>	<b>01</b>	<b>16.66%</b>
<b>Oxacillin</b>	<b>04</b>	<b>66.70%</b>	<b>02</b>	<b>33.30%</b>
<b>Gentamicin</b>	<b>04</b>	<b>66.70%</b>	<b>02</b>	<b>33.30%</b>
<b>Ciprofloxacin</b>	<b>05</b>	<b>83.33%%</b>	<b>01</b>	<b>16.66%</b>
<b>Levofloxacin</b>	<b>02</b>	<b>33.30%</b>	<b>04</b>	<b>66.70%</b>
<b>Inducible Clindamycin Resistance</b>	<b>05</b>	<b>83.33%%</b>	<b>01</b>	<b>16.66%</b>
<b>Erythromycin</b>	<b>02</b>	<b>33.30%</b>	<b>04</b>	<b>66.70%</b>
<b>Co-trimoxazole</b> <b>[Trimethoprim+ Sulphamethoxale]</b>	<b>02</b>	<b>33.30%</b>	<b>04</b>	<b>66.70%</b>
<b>Clindamycin</b>	<b>02</b>	<b>33.30%</b>	<b>04</b>	<b>66.70%</b>
<b>Linezolid</b>	<b>01</b>	<b>16.66%</b>	<b>05</b>	<b>83.33%</b>
<b>Daptomycin</b>	<b>01</b>	<b>16.66%</b>	<b>05</b>	<b>83.33%</b>
<b>Teicoplanin</b>	<b>01</b>	<b>16.66%</b>	<b>05</b>	<b>83.33%</b>
<b>Vancomycin</b>	<b>01</b>	<b>16.66%</b>	<b>05</b>	<b>83.33%</b>
<b>Tetracycline</b>	<b>02</b>	<b>33.30%</b>	<b>04</b>	<b>66.70%</b>
<b>Tigecycline</b>	<b>01</b>	<b>16.66%</b>	<b>05</b>	<b>83.33%</b>
<b>Nitrofurantoin</b>	<b>02</b>	<b>33.30%</b>	<b>04</b>	<b>66.70%</b>
<b>Rifampicin</b>	<b>02</b>	<b>33.30%</b>	<b>04</b>	<b>66.70%</b>



**Table: 9. Percentage of Resistant & Susceptibility of isolated *Pseudomonas aerugenosa* to tested use of antibiotics in PUS/WOUND Culture [<18 Years]:**

**Total Isolates: 02**

<b>Antibiotics</b>	<b>R (No.)</b>	<b>R (%)</b>	<b>S (No.)</b>	<b>S (%)</b>
<b>Piperacillin + Tazobactam</b>	<b>01</b>	<b>50.00%</b>	<b>01</b>	<b>50.00%</b>
<b>Ceftazidime</b>	<b>01</b>	<b>50.00%</b>	<b>01</b>	<b>50.00%</b>
<b>Cefoperazone+Subactam</b>	<b>01</b>	<b>50.00%</b>	<b>01</b>	<b>50.00%</b>
<b>Cefepime</b>	<b>01</b>	<b>50.00%</b>	<b>01</b>	<b>50.00%</b>
<b>Ciprofloxacin</b>	<b>01</b>	<b>50.00%</b>	<b>01</b>	<b>50.00%</b>
<b>Levofloxacin</b>	<b>01</b>	<b>50.00%</b>	<b>01</b>	<b>50.00%</b>
<b>Amikacin</b>	<b>01</b>	<b>50.00%</b>	<b>01</b>	<b>50.00%</b>
<b>Aztreonam</b>	<b>01</b>	<b>50.00%</b>	<b>01</b>	<b>50.00%</b>
<b>Imipenem</b>	<b>01</b>	<b>50.00%</b>	<b>01</b>	<b>50.00%</b>
<b>Meropenem</b>	<b>01</b>	<b>50.00%</b>	<b>01</b>	<b>50.00%</b>
<b>Colistin</b>	<b>01</b>	<b>50.00%</b>	<b>01</b>	<b>50.00%</b>

**Table: 10. Percentage of Resistant & Susceptibility of isolated *Pseudomonas aeruginosa* to tested use of antibiotics in PUS/WOUND Culture [ >18-64 Years]:**

**Total Isolates: 16**

<b>Antibiotics</b>	<b>R (No.)</b>	<b>R (%)</b>	<b>S (No.)</b>	<b>S (%)</b>
<b>Piperacillin + Tazobactam</b>	<b>04</b>	<b>25.00%</b>	<b>12</b>	<b>75.00%</b>
<b>Ceftazidime</b>	<b>04</b>	<b>25.00%</b>	<b>12</b>	<b>75.00%</b>
<b>Cefoperazone+Sulbactam</b>	<b>04</b>	<b>25.00%</b>	<b>12</b>	<b>75.00%</b>
<b>Cefepime</b>	<b>04</b>	<b>25.00%</b>	<b>12</b>	<b>75.00%</b>
<b>Ciprofloxacin</b>	<b>06</b>	<b>37.50%</b>	<b>10</b>	<b>62.50%</b>
<b>Levofloxacin</b>	<b>04</b>	<b>25.00%</b>	<b>12</b>	<b>75.00%</b>
<b>Amikacin</b>	<b>04</b>	<b>25.00%</b>	<b>12</b>	<b>75.00%</b>
<b>Imipenem</b>	<b>06</b>	<b>37.50%</b>	<b>10</b>	<b>62.50%</b>
<b>Aztreonem</b>	<b>12</b>	<b>75.00%</b>	<b>04</b>	<b>25.00%</b>
<b>Meropenem</b>	<b>04</b>	<b>25.00%</b>	<b>12</b>	<b>75.00%</b>
<b>Colistin</b>	<b>15</b>	<b>93.75%</b>	<b>01</b>	<b>6.25%</b>
<b>Cotrimoxazole</b>	<b>10</b>	<b>62.50%</b>	<b>06</b>	<b>37.50%</b>
<b>Trimethoprim+ Sulphamethoxale]</b>				

**Table: 11. Percentage of Resistant & Susceptibility of isolated *Pseudomonas aeruginosa* to tested use of antibiotics in PUS/WOUND Culture [>64 Years]:**

**Total Isolates: 06**

<b>Antibiotics</b>	<b>R (No.)</b>	<b>R (%)</b>	<b>S (No.)</b>	<b>S (%)</b>
<b>Piperacillin + Tazobactam</b>	<b>02</b>	<b>33.30%</b>	<b>04</b>	<b>66.70%</b>
<b>Ceftazidime</b>	<b>02</b>	<b>33.30%</b>	<b>04</b>	<b>66.70%</b>
<b>Cefoperazone+Sulbactam</b>	<b>02</b>	<b>33.30%</b>	<b>04</b>	<b>66.70%</b>
<b>Cefepime</b>	<b>04</b>	<b>66.70%</b>	<b>02</b>	<b>33.30%</b>
<b>Ciprofloxacin</b>	<b>04</b>	<b>66.70%</b>	<b>02</b>	<b>33.30%</b>
<b>Levofloxacin</b>	<b>04</b>	<b>66.70%</b>	<b>02</b>	<b>33.30%</b>
<b>Amikacin</b>	<b>02</b>	<b>33.30%</b>	<b>04</b>	<b>66.70%</b>
<b>Imipenem</b>	<b>02</b>	<b>33.30%</b>	<b>04</b>	<b>66.70%</b>
<b>Aztreonam</b>	<b>05</b>	<b>83.33%</b>	<b>01</b>	<b>16.66%</b>
<b>Meropenem</b>	<b>02</b>	<b>33.3%</b>	<b>04</b>	<b>66.7%</b>
<b>Colistin</b>	<b>04</b>	<b>66.70%</b>	<b>02</b>	<b>33.30%</b>
<b>Gentamicin</b>	<b>03</b>	<b>50.00%</b>	<b>03</b>	<b>50.00%</b>



#### IV. DISCUSSION

Despite adherence to basic pus/wound care principles, many patients still develop infections that necessitate accurate identification of causative organisms for appropriate management. The spectrum of isolated organisms and their antimicrobial susceptibility patterns often vary between hospitals, highlighting a dynamic trend. The development and spread of antibiotic resistance can be mitigated through the judicious use of antimicrobials, stringent infection control practices, and ongoing surveillance efforts.

Surgical site infections continue to pose a significant challenge in hospital settings, particularly in surgical wards where invasive procedures and prolonged hospital stays increase the risk of infection. This high isolation rate underscores the importance of microbiological evaluation of pus samples for targeted antimicrobial therapy.

*Klebsiella pneumoniae*, *Pseudomonas aeruginosa*, and *Staphylococcus aureus* are the primary pathogens isolated from pus and wound swabs, frequently showing high resistance to common antibiotics, including amoxicillin/clavulanic acid, 3rd generation cephalosporins, and high rates of MRSA/ORSA. *Pseudomonas aeruginosa* and *Staphylococcus aureus* are often the most prevalent, with *Klebsiella pneumoniae* dominating in surgical wards.

In this study *Klebsiella pneumoniae* in all age group shows Resistant in Cephalosporins group, Fluoroquinolones group, Penicillin group. Highly sensitive rate of Antibiotics is Tigecycline. Colistin highly Resistant antibiotics in *Klebsiella pneumoniae* and *Pseudomonas aeruginosa* in all age group. Piperacillin/Tazobactam and Cefoperazone/Sulbactam both are good response in *Pseudomonas aeruginosa* in all age group. Cefoperazone/Sulbactam also good performance in *Klebsiella pneumoniae* in all age group. In our study in all age group approx. 48.1% Oxacillin Resistant (ORSA) *Staphylococcus aureus* found. This findings is very important for our this clinical study. Linezolid and Tigecycline both are highly sensitive in *Staphylococcus aureus* in all age group.

This study provides the evidence of high prevalence of antibiotic resistant bacteria in pus samples of patients collected. The prevalence and antibiotics resistance patterns of pyogenic bacterial isolates usually exhibit variability according to geographic areas and climate conditions. Existence of high drug resistance to multiple antibiotics in *Staphylococcus aureus*, *Klebsiella pneumoniae*, and *Pseudomonas aeruginosa* isolates from pus samples.

In this study and several other related reports points towards negligence on patients part, incomplete treatment schedules, antibiotics misuse, self-prescription, misprescription, lack of regional antibiogram data, and limited knowledge about multidrug-resistant isolates and antimicrobial resistance among clinicians. Updated knowledge of antimicrobial susceptibility profiles of clinical isolates will not only assist in designing the most appropriate dose-regimen and treatment schedule against wound infections but also help in curbing the alarmingly expanding menace of drug resistance.

#### V. CONCLUSION

In conclusion, pyogenic wound infections were found prevalent in *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*. Bacterial isolates exhibited high to moderate levels of resistance against different classes of antibiotics. The susceptibility data from this report may be worth consideration while implementing empiric treatment strategies for pyogenic infections. At the same time, strict health policies should also be implemented to regulate the purchase and prescription and restrict the unsupervised antibiotic use as well as continuous monitoring and reporting antibiotic resistance.

This study indicates that treatment options for resistant bacteria pathogens are notably limited. Therefore, early detection and the judicious use of appropriate antibiotics are critical for managing and preventing the emergence and spread of multidrugresistant organisms.

Studies conclude that *Klebsiella pneumoniae*, *Pseudomonas aeruginosa*, and *Staphylococcus aureus* are highly prevalent in pus and wound swabs, frequently exhibiting multidrug resistance (MDR). High resistance is observed against commonly used antibiotics (e.g., amoxicillin/clavulanic acid, cephalosporins), while carbapenems, vancomycin, and linezolid often show higher efficacy.

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